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## Formation of Hydroxyapatite Coating using Novel Chemo-Biomimetic Method

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#### **Abstract**

A novel chemo-biomimetic method was developed to deposit hydroxyapatite (HA), simulating the porous nano-scale structure and chemical composition of natural bone. Electrochemical activation in NaOH solution, a prerequisite process to heterogeneously nucleate hydroxyapatite in this investigation, creates nano-scale porous structure on the surface of Ti6Al4V alloy. XPS analysis confirmed that the surface of activated Ti6Al4V substrate converted to TiO<sub>2</sub> during activation, existing in the form of hydrated TiO<sub>2</sub>. Benefited from the biocompatible top-layer of hydrated TiO<sub>2</sub> and the favorable alkaline surface chemistry created through the electrochemical activation, the HA coating nucleates heterogeneously and grows continuously on the activated substrate resembling the nano-scale porous bone-like structure. The coating was characterized using XRD, SEM/FESEM/EDX, TEM and FTIR, and was confirmed as pure hydroxyapatite. A coating thickness of 50μm was achieved, which is preferable and acceptable for medical implant application to promote bone ingrowth, thus enhancing fixation and biocompatibility of implant surface.

**Key Words:** Chemo-biomimetic, hydroxyapatite, Ti6Al4V alloy, electrochemical activation.

#### 1. Introduction

The prevalence of thigh pain (reported of up to 26% [1, 2]) after total hip replacement remains an unacceptable complaint by patients. The cause is probably due to a number of factors including unstable fibrous fixation, the mismatch between the moduli of elasticity for the prosthesis material and bone, fatigue fracture, relative motion across the interface between the implants and the surrounding tissues due to excessive stress at the implant/body tissue interfaces, and endosteal irritation [3-4]. Furthermore, materials implanted *in vivo* essentially have direct contact with human body through the interface between the implant surface and bones, tissues and extracellular body fluids. The surface of the implant therefore plays a very important role related to surface chemistry, topography and micro/nano structure, and tribological properties. This requires a surface modification to promote the ingrowth of human bone into the implanted biomaterials, more preferentially to form a natural and smooth connection between bone and implant.

Titanium and titanium alloys, bioinert materials due to a stable oxide layer at the surface [5-7], have been extensively investigated and applied as an implanting material due to their combined properties of corrosion resistance and biocompatibility with mechanical performance [8]. But these artificial materials implanted are generally encapsulated by a soft fibrous tissue, leading to isolation from the surrounding bone and a weak bonding between implant and bone [8, 9]. The calcium phosphate ceramics, bioactive materials, on the other hand can promote osteoconductivity and create a direct chemical bonding between implant and surrounding bone [8]. Much attention has therefore been focused on ceramics which resemble the mineral phase in bone tissues, *i.e.* hydroxyapatite, octacalcium phosphate and tricalcium phosphate [8, 10-12]. To take advantage of the excellent characteristics of Ti-alloys and to avoid the isolating layer of soft tissue around the Ti implants, the growth of apatite coating onto the Ti-alloy is currently being extensively investigated [9, 12-19].

However, most of the techniques for apatite coating are line-of-sight dependant and/or involves high temperature. It is a tremendous challenge for any process that is line-of-sight in nature to produce uniform coating, particularly on complex geometries; As for processes that rely on high temperature, they cause decomposition of HA which leads to the formation of impurities such as tetracalcium phosphate ( $Ca_4P_2O_9$ ), amorphous calcium phosphate,  $\alpha$ - and/or  $\beta$ -tricalcium phosphate ( $Ca_3(PO_4)_2$ ). These impurities are relatively unstable in the body fluids as compared to HA and the uneven surface formed due to the selective dissolution of these impurities may result in an accelerated wear caused by the roughening of surface, and the debris will, in turn, make the wear a more severe issue.

Biomimetic [14-16] and chemical processes [20, 21] are neither line-of-sight dependant nor involves high temperature operation. Biomimetic coating is an approach that consists of immersion of metal implants in simulated body fluids (SBF) at a physiologic temperature and pH. HA coating, the major component of bone, grows in a way similar to the natural bone growth in human body. This process produces HA coating with desirable properties such as high purity and bioactivity. However, achieving a reasonable coating thickness through biomimetic method for practical applications takes a long time. As a result of the low solubility product of HA and the limited concentration range for the metastable phase, this operation is extremely difficult and might lead to local precipitation or uneven coatings. Such an intricate and long process can hardly be tolerated in the prostheses coating industry. Chemical processes produce coatings at low temperature and is line-of-sight independent. It is a process to produce HA coating based simply on chemical reaction. Theoretically, this process can produce uniform coating of unlimited thickness on complex shapes. The deposition rate of chemical HA coating is significantly higher than the biomimetic process due to significantly higher and controllable solution composition. Unfortunately, little is known about its chemical reaction kinetics and the process is used mainly for producing HA powders.

In order to combine the advantages of biomimetic and chemical processes, extensive research was conducted in our laboratory to develop a novel process for coating of

hydroxyapatite on Ti6Al4V alloy for orthopaedic applications. The novel approach presented in this communication is termed as chemo-biomimetic HA coating process for two reasons: 1) the deposition is based on the conventional biomimetic process simulating the natural bone growth, and 2) chemical modifications were conducted to the substrate surface chemistry prior to HA deposition as well as to the composition of deposition solutions, more specifically, increased calcium and phosphate concentrations. Both modifications are designed to accelerate the deposition process.

#### 2. Experimental Details

#### 2.1 Materials and Preparation of Samples

A Ti6Al4V-ELI alloy (RMI Titanium Company, Mississauga, Canada) with a chemical composition (wt. %) of 0.12 O, 0.02 C, 0.08 N, 0.213 Fe, 6.16 Al, 3.92 V and balance Ti was chosen as the testing material. The specimens ( $10 \times 10 \text{mm}^2$ ) were sheared from a rolled plate (1.7mm thick), mechanically polished using SiC paper #400 and #600, and  $9 \mu \text{m} \text{Al}_2\text{O}_3$  paper.

A Ti wire was spot-welded to the polished sample for handling, and the polished samples were ultrasonically cleaned in acetone, ethanol and deionized water for 10 minutes each, separately. The clean specimens were then etched in a 0.2 vol.% HF + 0.4 vol.% HNO<sub>3</sub> acid solution for 10 minutes, rinsed thoroughly with deionized water, and subjected to ultrasonic cleaning for 10 minutes in deionized water and dried at room temperature. The specimens were ready for the further electrochemical activation treatment.

#### 2.2 Electrochemical Activation Treatment of Ti6Al4V Substrates

The electrochemical activation treatment of Ti6Al4V alloy prior to HA deposition was carried out at room temperature in a 10M NaOH solution for 30 minutes. A constant voltage of 10 volts was applied using HP E3610A DC power supply on the cleaned and etched samples (as working electrode). Two identical Ti6Al4V alloy plates (30×30mm²), equally distanced from the working electrode, were used as counter electrodes. After the electrochemical activation, the specimen was thoroughly rinsed using deionized water for subsequent apatite deposition.

#### 2.3 The Chemo-Biomimetic Method for Coating Hydroxyapatite

The apatite coating was formed in a solution with enhanced  $\text{Ca}^{2+}$  and  $\text{HPO}_4^{2-}$  concentrations by a factor of five, 5(Ca&P), shown in Table 1, as compared with the conventionally used simulated body fluid (SBF) and blood plasma. All the chemicals used for the solution are reagent grade, including NaCl (Fluka,  $\geq 99.5\%$ ), NaHCO<sub>3</sub> (Sigma-Aldrich,  $\geq 99.7\%$ ), KCl (Fisher,  $\geq 99.6\%$ ), Na<sub>2</sub>HPO<sub>4</sub> (Sigma-Aldrich,  $\geq 99.0\%$ ), MgCl<sub>2</sub>.6H<sub>2</sub>O (Sigma-Aldrich,  $\geq 99.0\%$ ), CaCl<sub>2</sub>.6H<sub>2</sub>O (Fisher,  $\geq 99.5\%$ ), Na<sub>2</sub>SO<sub>4</sub> (Anachemia,  $\geq 99.0\%$ ). The solution pH was adjusted to 6.50 using HCl and TRIS (trishydroxymethyl aminomethan).

Table 1 Chemical composition (mM) of SBF, blood plasma and 5(Ca&P) solution

	Na <sup>+</sup>	K <sup>+</sup>	Mg <sup>2+</sup>	Ca <sup>2+</sup>	Cl <sup>-</sup>	HCO <sub>3</sub>	HPO <sub>4</sub> <sup>2-</sup>	SO <sub>4</sub> <sup>2-</sup>
5(Ca&P)	142.0	5.0	1.5	12.5	159.0	4.2	5.0	0.5
SBF	142.0	5.0	1.5	2.5	148.8	4.2	1.0	0.5
Blood plasma	142.0	5.0	1.5	2.5	103.0	27.0	1.0	0.5

The coating process was conducted in 250ml beakers with 175ml solution, and the beakers were kept in a water bath at 37°C. The coating process was allowed to proceed for 30 minutes before new solution was replenished and this replenishing was conducted at 30 minute intervals thereafter until completion of deposition. Another coating process was allowed to proceed continuously for 5 hours, without replenishment, to identify the effects of replenishment.

#### 2.4 Characterization of Formed Coating

The surface morphologies of hydroxyapatite coating on Ti6Al4V alloy were observed using Hitachi S-3500 SEM/EDX and Hitachi S-4500 field emission SEM (FESEM) equipped with an EDAXTM EDX system.

The nano grain size of formed hydroxyapatite coating was characterized using Philips CM20 Transmission Electron Microscope (TEM). The coating was scratched off the Ti6Al4V alloy substrate, dispensed in Ethanol, attached on a copper screen mesh for TEM analysis.

The coating was also scratched off the coated substrates and enough amount of powder was collected for XRD analysis. XRD analysis was carried out using a Bruker D8 diffractometer with Cuk $\alpha$  radiation. The scan was conducted from 10 to 70° at a step of 0.02°.

The coatings were also analyzed using Perkin Elmer FTIR (Fourier Transform Infrared Spectroscopy) Instrument (Spectrum BX), with the scanning range set between 4000 and 400 cm<sup>-1</sup> for 216 scans for each sample.

X-ray photoelectron spectroscopy (XPS) was carried out on the activated Ti6Al4V alloy substrate with a Kratos AXIS Ultra spectrometer. XPS is a very surface sensitive analytical technique and, as such, provides elemental and chemical state data from the outer 5-10 nm of a surface. For this work, one set of analyses was carried out on each sample using a monochromatized Al K $\alpha$  X-ray source. The analysis area was  $\sim 700$  x 300  $\mu$ m in size. Survey (pass energy 160 eV) and high-resolution spectra (pass energy 20 eV) for Ti 2p, C 1s, and O 1s peaks were obtained for each sample.

#### 3. Results and Discussion

#### 3.1 Activated Ti6Al4V Alloy Substrate

The XPS analysis shows that the surface of the activated Ti6Al4V substrate was completely converted to TiO<sub>2</sub> during activation (Fig. 1). The Ar<sup>+</sup> sputtering cleaned Ti6Al4V specimen (Fig. 1a) showed two peaks at the binding energies of 460.0eV and 453.9eV, attributable to Ti 2p1/2 and Ti 2p3/2 peaks, respectively. High resolution XPS spectrum of Ti 2p peaks (Fig. 1b) obtained for the electrochemically activated specimen showed no Ti 2p metal components, suggesting the formation of a thicker oxide layer on the surface (>5-10 nm). Both Ti 2p peaks corresponded to Ti<sup>4+</sup> valence state, indicating that suboxides such as TiO, Ti<sub>2</sub>O<sub>3</sub> or Ti<sub>3</sub>O<sub>5</sub> were not present on the oxide surface. The O 1s peak (Fig. 1c) was fitted with three different components, lattice oxide (O<sup>2-</sup>), hydroxide (physisorbed -OH), and water (chemisorbed H<sub>2</sub>O). The specimen showed a major peak at 530.5  $\pm$  0.3 eV, which is attributed to lattice oxide. Minor O 1s peaks at higher binding energies are attributable to hydroxides (532.0 eV) and chemisorbed H<sub>2</sub>O (533.2 eV).

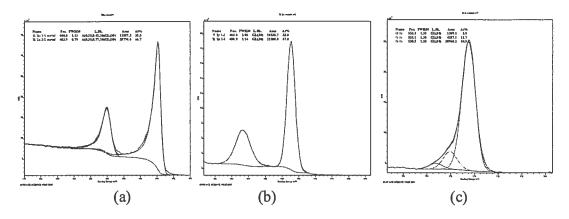


Fig.1 High resolution XPS spectrum of Ti 2p for (a) Ar<sup>+</sup> sputtered and (b) electrochemically activated Ti6Al4V alloys, and of O 1s for (c) electrochemically activated Ti6Al4V alloy

The electrochemically activated surface was verified as alkaline chemistry by surface pH measurement using pH indicators, shown in Table 2. These measurements revealed that the surface pH for etched Ti6Al4V alloy was in the range of 5.2 to 7.4; and that for the electrochemically activated Ti6Al4V alloy was equal or greater than 8.2, which is favorable for the formation of hydroxyapaptite.

Table 2 Surface pH range of etched and electrochemically activated Ti6Al4V alloy

Indicators		Methyl orange	Bromocresol purple	m-Cresol purple	Phenolphthalein	
Properties of indicators	pH range	3.2 – 4.4	5.2 – 6.8	1.2 – 2.8; 7.4 – 9.0	8.2 – 10.0	
	Color change	Red to yellow	Yellow to purple	Red to yellow; yellow to purple	Color to pink	
Reactions of samples to Etched		To yellow	To purple	To yellow	No change	
indicators	Activated			To purple	To pink	

#### 3.2 Deposition of Hydroxyapatite Coating

#### 3.2.1 Continuous Coating Process

In this chemo-biomimetic method, hydroxyapatite coating heterogeneously deposits on the Ti6Al4V alloy substrate due to both the nano-scale porous bone-like structure (With pore diameter of about 100nm) (Fig. 2a) with top-layer of TiO<sub>2</sub> and favorable surface alkaline chemistry created through electrochemical activation in the NaOH solution. The formed coating, especially the initial layer of the coating (Fig. 2b) has similar surface structure as the NaOH-activated substrate but with a larger pore diameter (about 1.0μm) indicating a good potential for retaining proteins and antibiotics that are conducive to post surgical healing. Cross-section of the initial coating (Fig. 2c) shows that the initial deposition forms a fine layer of coating with porous morphology, tightly attached to the substrate without coating loss along the edge of Ti6Al4V alloy substrate during the mechanical breakage of the sample.

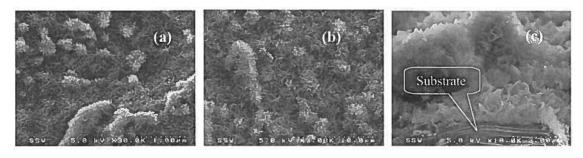


Fig. 2 FESEM presentations of (a) the surface of an activated Ti6Al4V alloy substrate, (b) surface morphology and (c) cross-section of the initial HA coating at 0.5 hour of immersion in the solution

The coating process on the activated Ti6Al4V alloy proceeds in the sequence of nucleation stage and growth stage. In the nucleation stage (Fig. 3a) apatite coating initially deposits on the activated surface through heterogeneous nucleation with spherical extrusions. The coating resembles porous bone-like structure and covers the whole surface of the substrate, even though only a short period of deposition is applied. At the growth stage, coating starts to grow in the valleys around the spherical features to create a leveled surface (Fig. 3b), which eventually demonstrates crystal-like morphologies at the fifth hour of deposition (Fig. 3c). It is shown from cross-section of the coating (Fig. 3d) that the continuous growth still creates fine porous structure throughout the coating. The cross-section of the coated Ti6Al4V was prepared by mechanically breaking the samples using a vise. It can be clearly seen from the fractured surface that the even though a breakage is observable inside the coating, the initial layer of deposit still well adheres to the substrate, indicating a good interface bonding.

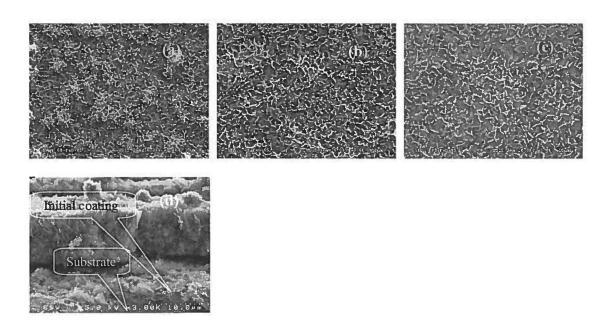


Fig. 3 Surface morphologies of apatite coating formed on activated Ti6Al4V alloy in the initial stage of (a) 0.5 hour and the growth stage of (b) 2 hours and (c) 5 hours; (d) cross-section of coating at 5 hours

Fig. 4 shows calculated coating thickness and corresponding pH variation of bulk solution. The coating thickness was calculated from the weight gain of deposit on the substrate based on the full density of hydroxyapatite (3.16g/cm³). During the initial heterogeneous nucleation, the pH of the bulk solution is measured to be relatively unchanged. Accordingly, a relatively stable coating growth is maintained. At the growth stage, slightly increased pH at the second hour corresponds to a significant increase of coating thickness on the substrate. With the relatively high deposition rate of HA, chemicals in the solution are quickly depleted, leading to a significant reduction of pH in the solution and a reduced deposition rate. Replenishing of the solution at certain intervals needs to be addressed.

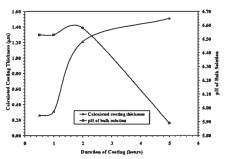


Fig. 4 Plot of calculated coating thickness as function of corresponding pH of bulk solution

#### 3.2.2 Replenishing Solutions during Coating Processes

The purpose of replenishing solution is to investigate the deposition and growth of coating under "constant" concentration and pH of the solution, and maintain the deposition rate. The process was carried out by replenishing solution every half an hour with fresh solution, and necessary analysis was conducted on the coated samples retrieved at 0.5, 1, 2, 3, 4 and 5 hours. The morphologies of coated Ti6Al4V substrates at different duration show that all coatings are bone-like structure and cover the whole surface of the Ti6Al4V alloy substrate. Coating morphology at half an hour (Fig. 5a) demonstrates that HA coating grows in a spherical shape across the surface, resulting in an uneven coating surface. Gradually the valleys around the spherical features are covered (Fig. 5b), resulting in a ball-like and relatively even coating surface with continued deposition. From the 4th hour (Fig. 5c), the coating grows to certain thickness and starts to crack due to the accumulation of internal stress. Compared with the continuous coating process, the coatings in both processes are of similar bone-like morphology for the first two hours, while the replenishing process maintains the dense and bone-like structure for the whole duration of coating with smaller pore size. The cross-section observation confirms that the coating is uniform on the edge, corner and the surface (Fig. 5d), attributing to the non line-of-sight nature of this method.

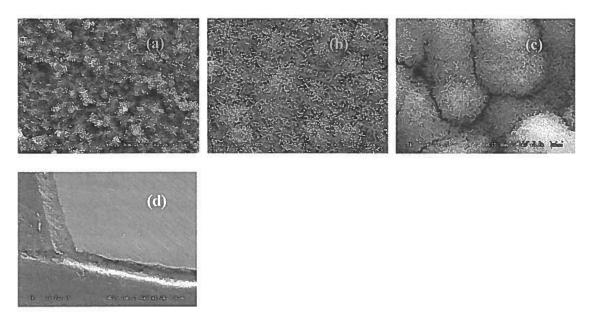


Fig. 5 Surface morphologies of HA coating at (a) 0.5, (b) 2 and (c) 4 hours and (d) cross-section of 4 hours of coating duration with replenishing solution every half an hour

A linear relationship is observed (Fig. 6a) between the calculated coating thickness (from weight gain basing on full density of hydroxyapatite) and the duration of coating. Comparing with the coating thickness of one hour interval of replenishing and without replenishing for the 5-hour coating process, replenishing solutions significantly increases the coating growth on the substrate. In other words, replenishing enables a steady growth of HA. It ought to be mentioned that the calculated coating thickness does not represent the actual coating thickness due to the large amount of porosities in the coating. The actual coating thickness was measured from the cross-section of the coated samples using

SEM (Fig. 6b). Compared with the calculated coating thickness, the actual coating thickness is much greater and also linearly related to the duration of deposition.

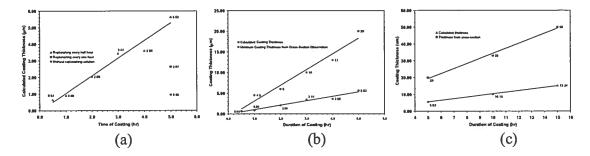


Fig. 6 (a) Effect of replenishing solution on the calculated coating thickness, (b) comparison between the calculated and actual coating thickness for (b) short-term (5 hours) and (c) long-term (15 hours) coating processes

To further confirm the linear growth of coating and the reliability of this coating process, long term deposition was carried out. The results confirm that a linear relationship between the coating thickness (actual) and the duration of coating process still exists (Fig. 6c). Coating thickness reaches 50µm within 15 hours, which is an acceptable thickness of hydroxyapatite coating for medical application. EDX analysis was conducted to define boundaries of the coating, and confirms that the thickness is around about 33µm for the initial 10 hours (Fig. 7a) and 50µm for the 15 hours (Fig. 7b) of deposition.

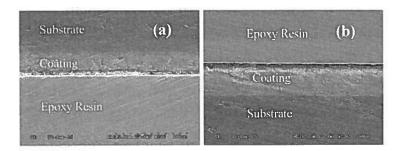
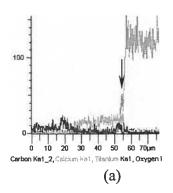


Fig. 7 Cross-section of coated Ti6Al4V alloy at (b) 10 and (c) 15 hours of coating process

#### 3.3 Characterization of Formed Coating

#### 3.3.1 EDX Analysis of Coating

EDX line analysis conducted from the coating to the substrate (Fig. 8a) confirms that there is an initial layer with diffusion of Ti and Ca (about  $2-3\mu m$ ), indicated by an arrow at the interface of substrate and the HA coating. This is possibly the result of the preferential reaction between  $TiO_2$  and  $Ca^{2+}$  at the initial stage. The reaction possibly proceeds inside the pores (created by electrochemical activation) through the diffusion of  $Ca^{2+}$  into the pores, rather than the diffusion of Ti out of substrate.



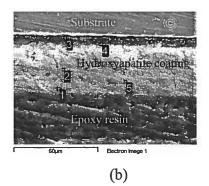


Fig. 8 (a) EDX line analysis to confirm the initial diffusion layer and (b) EDX point analysis of the coating

During the electrochemical activation process in the alkaline solution, the naturally formed thin titanium oxide is easily dissolvable due to its reaction with OH<sup>-</sup> to form HTiO<sub>3</sub>. The subsequently exposed Ti is electrochemically oxidized in the alkaline solution possibly according to the following reaction (1):

$$Ti + 3OH^{-} - 3e \rightarrow TiO_{2}.H_{2}O + \frac{1}{2}H_{2} \uparrow \tag{1}$$

Gas evolution was observed on the Ti6Al4V substrate, possibly the result of hydrogen formation. The XPS analysis (Fig. 1) shows that the O is mainly in the form of lattice oxide (O<sup>2-</sup>) with small amount of hydroxide (physisorbed OH<sup>-</sup>) and water (chemisorbed H<sub>2</sub>O), thus the surface of activated substrate is dominated by hydrated TiO<sub>2</sub> and possibly Ti-OH<sup>-</sup> hydrogel layer. The nano-scale pores formed during activation process may also demonstrate a capillary effect, which absorbs and retains the NaOH solution in the pores, resulting in a higher local pH on the surface. In addition to the biocompatible TiO<sub>2</sub>, the alkaline environment on the surface from both the capillary effect and Ti-OH<sup>-</sup> layer is beneficial for the heterogeneous nucleation of hydroxyapatite on the surface, according to the following reaction (2):

$$10Ca^{2+} + 6PO_4^{3-} + 2OH \leftrightarrow Ca_{10}(PO_4)_6(OH)_2 \tag{2}$$

One of the main characteristics of metal oxides in aqueous solution is their point of zero charge (PZC), which represents the pH value of aqueous solution at which an immersed oxide surface has zero net charge. If the solution pH is greater than the pH at PZC of the metal oxide, the metal oxide is negatively charged; on the contrary, if the solution pH is lower than the pH at PZC of metal oxide, the metal oxide is positively charged. The charged surface and surrounding solution form a thin double layer, to which oppositely charged ions are attracted and initial reaction proceeds. The PZC of the TiO<sub>2</sub> is reported to be in the range of 5.5 to 6.0 [22], resulting in a negatively charged surface in the coating solution (The pH was measured of 6.50 in this investigation). When the positively charged Ca<sup>2+</sup> is attracted to the surface of Ti6Al4V alloy, it reacts with the TiO<sub>2</sub> to form CaTiO<sub>3</sub>, the PZC of which was reported as 8.1 [23], causing the CaTiO<sub>3</sub>—

covered surface to be positively charged and the PO<sub>4</sub><sup>3-</sup> attracted to the surface to participate in further reactions, leading to the formation of calcium phosphate.

The results of EDX point analysis (Fig. 8b) on the hydroxyapatite coating and corresponding ratios of Ca/P are listed in Table 3. Measurements 3 and 4 close to the substrate show the coexistence of Ca, Ti, V or P, indicating a preferential formation of Ca on the Ti6Al4V surface at the initial stage of deposition (no phosphor was detected in measurement 3). The phosphor then participates in subsequent deposition, as is evident in measurement 4, away from the substrate surface. In summary, EDX point analysis confirms the existence of initial diffusion layer and the sequential participation of Ca<sup>2+</sup> and PO<sub>4</sub><sup>3-</sup> in the formation of HA coating.

With the formation of calcium phosphate apatite on the surface, the super-saturation of simulated physiological solution may promote preferential nucleation of apatite on the already-formed apatite rather than in the solution, the formed hydroxyapatite acts as seeds to accelerate the formation of the hydroxyapatite [24, 25]. Thus the further formation of the apatite coating continues on the initial apatite layer by spontaneous growth consuming calcium and phosphate ions from the surrounding simulated body fluid. A theoretical analysis [26] also indicates that formation of hydroxyapatite exhibits a higher thermodynamic preference than that of octacalcium phosphate (OCP) and dicalcium phosphate (DCPD). Thus the further formation of the coating proceeds in the form of hydroxyapatite, rather than OCP or DCPD. This effect is evident by the eventual formation of hydroxyapatite shown by measurements 1, 2 and 5, which are away from the initial deposition layer and the substrate. The average of Ca/P ratios of these three measurements is 1.67, which is exactly the atomic ratio of Ca/P in stoichometric hydroxyapatite ( $Ca_{10}(PO_4)_6(OH)_2$ ).

Table 3 EDX analysis (atomic percentage) on the cross-section of coating

			(							
	Measurement	С	0	P	Cl	Ca	Ti	V	Ca/P	Average
Initial Layer	3	21.80				21.47	51.55	5.19	N/A	N/A
	4	32.71	25.72	5.25		21.47	13.68	1.23	4.08	
Bulk Coating	2	58.04	18.72	8.33	0.56	14.35			1.72	1.67
	5	64.56	18.11	6.36	0.36	10.59			1.67	
	1	67.77	16.43	5.81	0.52	9.47			1.63	

#### 3.3.2 FTIR, XRD and TEM Characterization of Coating

XRD analysis (Fig. 9) of a commercial crystallized hydroxyapatite powder (PENTAX Corporation, Tokyo, Japan) and the comparison with the powders of formed coating confirms that the formed coating is hydroxyapatite. The commercial HA XRD patterns (lower curve) match very well with the standard XRD pattern of HA in the JCPDF card, major patterns of the coating powders (upper curve) are shown and match with the commercial HA powders.

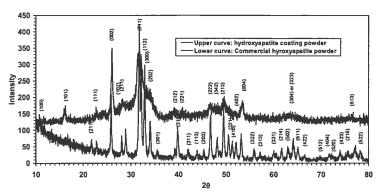


Fig. 9 XRD analysis of a commercial HA powder and powder of formed coating

Fig. 10 is the FTIR spectra at different duration of coating process. The spectra show the characteristic bands of hydroxyapatite along with additional bands of associated H<sub>2</sub>O. It is concluded that the formed apatite coating is hydroxyapatite, consistent with the XRD analysis.

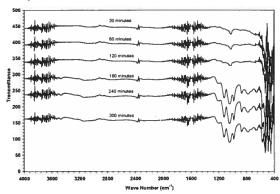


Fig. 10 FTIR spectra of coatings at different durations

The stretching vibration of  $PO_4^{3-}$  can be observed in the range of 1200 - 900 cm<sup>-1</sup>, which in this investigation located at 1119, 1048, 1036 and 979 cm<sup>-1</sup>; the deformation vibration of  $PO_4^{3-}$  are located at the bands of 603, 572, 471 and 422 cm<sup>-1</sup>; as for hydroxyl (OH<sup>-</sup>), the bands are located at 3458 and 617 cm<sup>-1</sup>. The fluctuations located in the range of 3900 – 3500 and 1900 – 1400 can be assigned to the associated  $H_2O$  in the coating, a possible result of long period exposure of porous HA in the atmosphere. The intensities of the hydroxyapatite bands are significantly increased with the increasing duration of coating.

The broad characteristic bands of HA illustrate amorphous or poorly crystallized phases of initially deposited HA during the first 120 minutes. The bands split after 180 minute deposition shows that the formed HA is gradually crystallized, but still not fully crystallized due to the relative broadness of the band peaks. The sharpness of bands, especially that of the 572 and 601 cm<sup>-1</sup> bands, indicates a well-crystallized HA.

Fig. 11a is the TEM diffraction patterns of the formed HA coating, showing that the formed HA coating has crystal structure and the measured d-spacings match with that of pure HA (corresponding d-spacing numbers for pure HA are 4.72, 3.51, 3.17, 2.81, 2.53, 2.30, 1.89, 1.59, 1.50, 1.43, 1.35). The formed HA coating has nano-scale grain size of

less than 20nm, the crystal planes are shown inside the grains and corresponding d-spacing can be measured (Fig. 11b). Morphologies of a single HA plate also show that the grain is less than 20nm in size (Fig. 11c).

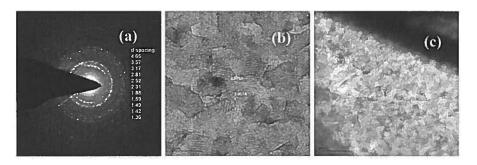


Fig. 11 TEM analysis of formed coating in terms of (a) diffraction pattern, (b) grin size and crystal planes, and (c) morphology of grains

#### **Conclusions**

A novel HA coating process is developed to deposit hydroxyapatite coating with porous bone-like structure on Ti6Al4V alloy. This method is capable of producing uniform deposit due to its non line-of-sight nature. The deposition proceeds through heterogeneous nucleation on the activated substrate due to the biocompatible TiO<sub>2</sub> and favorable surface chemistry created through the electrochemical activation in NaOH solution. Growth of the coating from a replenished solution is linearly related to the coating duration and reaches 50µm within 15 hours. The formed coating resembles the porous bone-like features of the electrochemically activated substrate, and is characterized as pure hydroxyapatite using XRD and FTIR analysis. Further, the deposit features nano-scale grain size.

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#### References

- 1. R. Bourne, C. Rorabec, J. Patterson and J. Guerin, "Tapered titanium cementless total hip replacements. A 10-13 year follow-up study". Clin Orthop, 393 (2001):112.
- 2. H. Kawamura, M. Dunbar, P. Murray, R. Bourne and C. Rorabeck, "The porous coated anatomic total hip replacement: A ten to fourteen year follow-up of cementless total hip arthroplasty". J Bone Joint Surg, 83A (2001):1333.

- 3. Campbell, A., Rorabeck, C., Bourne R., Chess D., Nott L., "Thigh pain after cementless hip arthroplasty. Annoyance or ill omen". J. Bone Joint Surg. Br. 74 (1993): 63-66.
- 4. Bourne R., Rorabeck C., Ghazal M., Lee M., "Pain in the thigh following total hip replacement with a porous-coated anatomic prosthessis for osteoarthritis. A five-year follow-up study". J. Bone Joint Surg. Am. 76 (1994):1464-1470.
- J. Lausmaa, B. Kasemo and H. Mattsson, "Surface spectroscopic characterization of Titanium implant materials", Applied Surface Science, Vol. 44 (1990), pp. 133-146.
- 6. E. McCafferty and J. P. Wightman, "An X-ray photoelectron spectroscopy sputter profile study of the native air-formed oxide film on titanium", Applied Surface Science, Vol. 143 (1999), pp. 92-100.
- 7. J. Pouilleau, D. Devilliers, F. Garrido, S. Durand-Vidal and E. Mahé, "Structure and composition of passive titanium oxide films", Materials Science and Engineering, Vol. B47 (1997), pp. 235-243.
- 8. D. M. Brunette, P. Tengvall, M. Textor and P. Thomsen, Titanium in Medicine, Springer-Verlag Berlin Heidelberg New York, USA, 2001.
- 9. H. M. Kim, F. Miyaji, T. Kokubo and T. nakamura, "Preparation of bioactive Ti and its alloys via simple chemical surface treatment", Journal of Biodical Materials Research, Vol. 32 (1996), pp. 409-417.
- 10. K. Hata and T. Kokubo, "Growth of a bonelike apatite layer on a substrate by biomimetic process", Journal of the American Ceramic Society, Vol.78 (1995), pp.1049-1053.
- 11. Y. Tang, Y. Liu, U. Sampathkumaran, M. Z. Hu, R. Wang and M. R. De Guire, "Particle growth and particle surface interaction of ceramic thin films", Solid State Ionics, Vol. 151 (2002), pp69-78.
- 12. Lee, T. M., B. C. Wang, Y. C. Yang, E. Chang, and C. Y. Yang. "Comparison of Plasma-Sprayed Hydroxyapatite Coatings and Hydroxyapatite/Tricalcium Phosphate Composite Coatings: In Vivo Study." Journal of Biomedical Materials Research (USA), 55, no. 3 (2001): 360-367.
- 13. Ding, S. J., C. P. Ju, and J. H. C. Lin. "Morphology and Immersion Behavior of Plasma-Sprayed Hydroxyapatite /Bioactive Glass Coatings." Journal of Materials Science: Materials in Medicine (USA), 11, no. 3 (2000): 183-90.
- S. Hayakawa and A. Osaka, "Biomimetic deposition of calcium phosphates on oxides soaked in a simulated body fluid", Journal of Non-Crystalline Solids, Vol. 263&264, (2000), pp. 409-415.
- 15. J. Wang, P. Layrolle, M. Stigter and K. de Groot, "Biomimetic and electrolytic calcium phosphate coatings on titanium alloy: physicochemical characteristics and cell attachment", Biomaterials, Vol. 25 (2004), pp.583-592.
- 16. L. Jonášová, F. A. Müller, A. Helebrant, J. Strnad and P. Greil, "Biomimetic apatite formation on chemically treated titanium", Biomaterials, Vol. 25 (2004), pp. 1187-1194.
- 17. A. Stoch, A. Brożek, G. Kmita, J. Stoch, W. Jastrzębski and A. Rakowska, "Electrophoretic coating of hydroxyapatite on titanium implants", Journal of Molecular Structure, Vol. 596 (2001), pp.191-200.

- 18. H. W. Kim, Y. H. Koh, L. H. Li, S. Lee and H. E. Kim, "Hydroxyapatite coating on titanium substrate with titania buffer layer processed by sol-gel method", Biomaterials, Vol. 25 (2004), pp. 2533-2538.
- 19. F. Barrere, M. M. E. Snel, C. A. van Blitterswijk, K. de Groot and P. Layrolle, "Nano-scale study of the nucleation and growth of calcium phosphate coating on titanium implants", Biomaterials, Vol. 25 (2004), pp. 2901-2910.
- 20. Tas, AC, "Synthesis of biomimetic Ca-hydroxyapatite powders at 37°C in synthetic body fluids", Biomaterials, Vol 21 (2000) no. 14 pp1429-1438.
- 21. Kong, LB; Ma, J; Boey, F. "Nanosized hydroxyapatite powders derived from coprecipitation process", Journal of Materials Science, Vol 37 (2002), no.6, pp. 1131-1134.
- 22. M. Kosmulski, "The pH-dependant surface charging and the points of zero charge", Journal of Collid and Interface Science, Vol. 253 (2002), pp 77-87.
- 23. T. hanawa, M. Kon, H. Doi, H. Ukai, K. Murakami, H. Hamanaka and K. Asaoka, "Amount of hydroxyal radical on calcium-ion-implanted titanium and point of zero charge of constituent oxide of the surface-modified layer", Journal of Materials Science: Materials in Medicine, Vol. 9 (1998), pp 89-92.
- 24. Roger I. martin and Paul W. Brown, "Aqueous formation of hydroxyapatite", Journal of Biomedical Materials Research, Vol 35 (1997), pp299-308.
- 25. Wenju Wu, Hengzhong Zhuang and George H. nancollas, "Heterogeneous nucleation of calcium phosphates on solid surfaces in aqueous solution", Journal of Biomedical Materials Research, Vol 35 (1997), pp93-99.
- 26. X. Lu, Y. Leng, "Theoretical analysis of calcium phosphate precipitation in the simulated body fluid", Biomaterials, Vol. 26 (2005), pp1097-1108.